

# Diagnostic accuracy of OLM's *CandID* PCR kit for detection of candidaemia: a preliminary evaluation

Jessica S. Price<sup>1</sup>, Melissa Fallon<sup>1</sup>, Gemma Johnson<sup>2</sup>, P. Lewis White<sup>1</sup>

1: Public Health Wales Mycology Reference Laboratory, Cardiff, United Kingdom

2: OLM Diagnostics, Braintree, United Kingdom



## Background

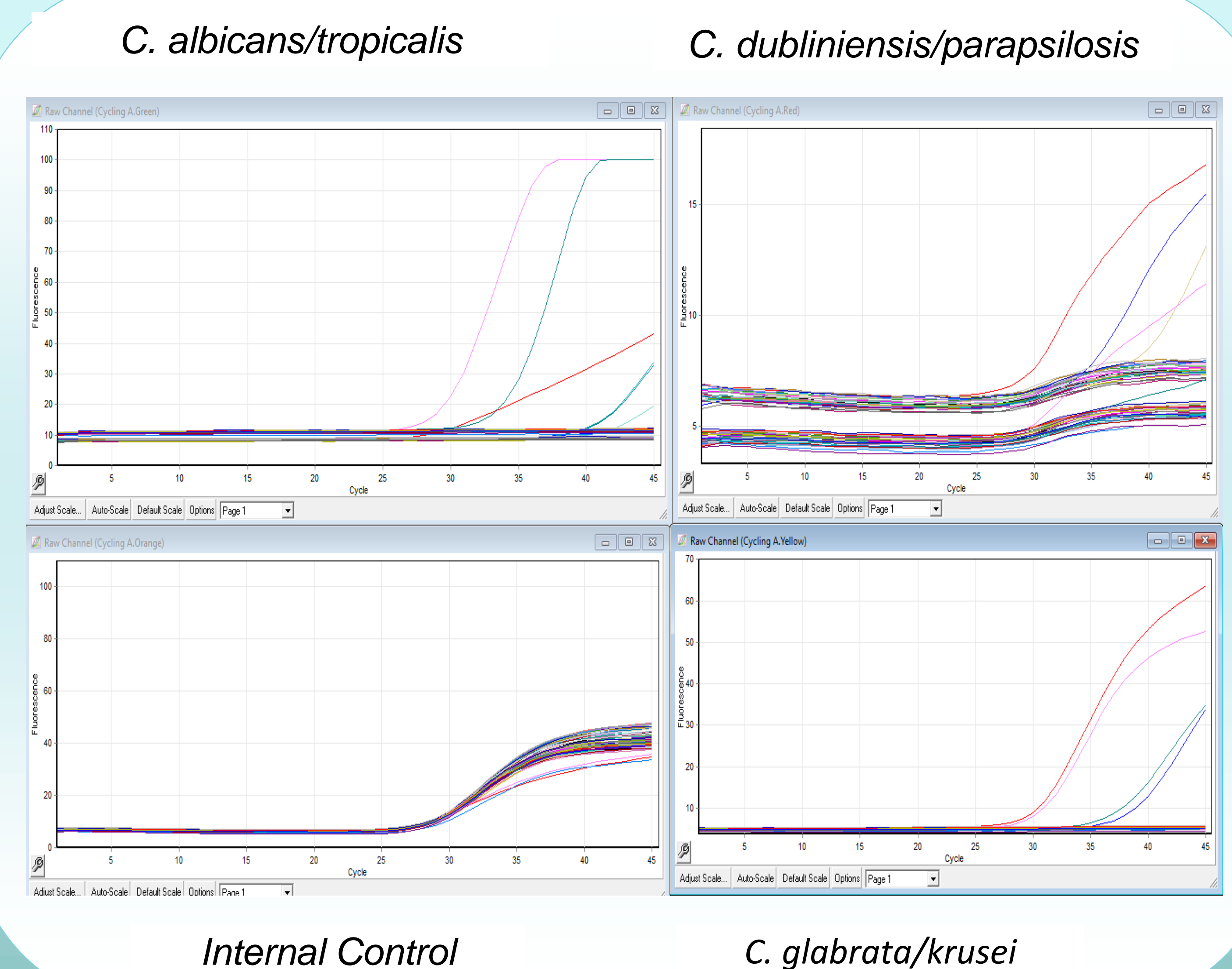
Treatment for candidaemia is time critical, with delayed treatment associated with higher hospital mortality. Culture-based tests lack sensitivity and prolong turnaround times, minimizing clinical utility. In this context, non-culture-based methods for the identification of *Candida*, such as PCR, represent a promising approach to allow rapid diagnosis and species identification, allowing the rapid initiation of species-oriented therapy. Our objective was to undertake an initial retrospective evaluation of the OLM Diagnostics *CandID* commercial PCR test for the diagnosis of invasive candida disease (IC).

## Materials/Methods

As part of routine testing across Wales, patients are prospectively screened for IC by Bruker Fungiplex *Candida* PCR and Fungitell (1-3)-β-D-Glucan (BDG). Samples from a 4-month period were retrospectively tested by the OLM *CandID* PCR assay. Initially, serum samples were selected based on a previous positive Fungiplex result, and required nucleic acid extraction of 0.5mL of serum (stored at -80°C) on the BioMerieux eMAG platform. Additional *CandID* testing was performed on randomly selected DNA eluates, previously extracted during routine testing and stored at 4°C prior to *CandID* testing performed on the Qiagen Rotorgene-Q, as per manufacturer's instructions. Performance parameters (Sensitivity, specificity Positive/negative likelihood ratios) of the *CandID* were calculated, with IC defined as follows: Candidaemia (n=10) - *Candida* recovered by blood culture; Probable IC (n=12) - Clinical risk factor for invasive candidiasis (e.g. surgery, gut disruption, COVID-19), plus evidence of *Candida* colonization at ≥2 non-contiguous anatomical sites and a positive serum (1-3)-β-D-Glucan; or recovery of *Candida* from a central venous catheter or deep wound and a positive serum (1-3)-β-D-Glucan. A control population of 45 patients with no evidence of IC was included. *Candida* PCR testing played no role in defining IC.

## Results

Nucleic Acid extraction through PCR amplification and result interpretation was completed in less than 4 hours. 74 serum samples were selected for testing (28 from 22 IC cases, 46 from 45 controls). Positivity rate for samples from cases was 75% (21/28, 95% CI: 57-87), significantly greater than the false positivity rate (6.5%, 3/46, 95% CI: 2.2-17.5) for control samples. Clinical performance is shown in the table. False positive results were associated with an identification of *C. parapsilosis* (n=1) and *C. krusei* (n=2). Using a threshold of <37.5% cycles to define PCR positivity increased specificity to 100%, with a slight reduction in sensitivity (77%). Species identification with *CandID* kits was 100% concordant with that obtained through culture identification (*C. albicans* (n=13), *C. glabrata* (n=4), *C. tropicalis* (n=1), *C. krusei* (n=1)). Two candidaemia cases falsely negative by PCR were also negative by BDG and routine PCR testing. One patient only had *Candida* recovered in 1/3 blood culture bottles, the other had received prior antifungal therapy.



Population	Sensitivity (%)	Specificity (%)	LR+tive	LR-tive	DOR
Candidaemia (n=10) vs No IC (n=45)	<b>80 (49-94)</b>	<b>93 (82-98)</b>	<b>11.43</b>	<b>0.22</b>	<b>53.14</b>
Probable IC (n=12) vs No IC (n=45)	<b>92 (65-99)</b>	<b>93 (82-98)</b>	<b>13.14</b>	<b>0.09</b>	<b>152.79</b>
Combined IC (n=22) vs No IC (n=45)	<b>86 (67-95)</b>	<b>93 (82-98)</b>	<b>12.3</b>	<b>0.15</b>	<b>81.61</b>

## Conclusions

*CandID* provides excellent performance and a rapid time-to-result, with accurate species identification that may prove beneficial in cases of suspected IC lacking candidaemia, by supporting clinical decision-making and avoiding treatment delay or inappropriate therapy choices for IC caused by non-*albicans* species. Retrospective data indicate the assay is useful for excluding but more so confirming IC. A prospective evaluation is required for confirmation.